EFFECT OF STORAGE CONTAINERS ON THE PERCENT INCIDENCE OF ASPERGILLUS SPECIES

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ABSTRACT:

The aim of present study is carried out to know the effect of storage containers on the percent incidence of *Aspergillus* species. Association of different species of *Aspergillus* with the seeds has been reported to cause decrease in percent germination of seeds, seed discoloration, heating and mustiness, loss of seed weight, seed rotting, biochemical changes, production of mycotoxins, etc. Storage containers play an important role in preventing or multiplying the original seed mycoflora in stored seeds of different crops. In order to confirm the seeds were stored in irrespective containers at room temperature for the period of three months. The seeds were analysed for their load of *Aspergillus* species by using Potato Dextrose Agar plates. Different *Aspergillus* species, isolated from seeds of jowar and groundnut. The seeds of both the crops (jowar and groundnut) showed maximum incidence of *Aspergillus* species that were stored in the tin boxes followed by polythene bags while, the seeds stored in cloth bags and gunny bags yielded less number of *Aspergillus* species.

KEY WORDS: Aspergillus species, Potato Dextrose Agar, stored containers, stored seeds

INTRODUCTION:

The isolation and identification of species of *Aspergillus* for which seeds of cereals, pulses, oil seeds, etc. were collected both from fields as well as from various market places. In all 14 species of *Aspergillus* with numerous strains were isolated by using both blotter and agar medium as recommended by ISTA (1966). The species like *A. flavus, A. niger, A, ruber, A. fumigatus* were found to be very common on all types of seeds used for isolation. While, species like *A. sulphureus, A. glaucus* and *A. japonicus* exhibited their association only with the seeds of some particular crops. The number of *Aspergillus* species on the seeds showed their association with the other moulds like species of *Alternaria, Dreschleria, Fusarium, Curvularia* and *Cladosporium* while, poor incidence of *Aspergillus* was noted on the seeds which showed of *Rhizoctonia, Syncephalostrum, Penicillum, Trichoderma* and Chaetomium. Seeds of most plant

species may be safely stored for several months by carful control of temperature and relative humidity (Lacerda *et al.*, 2003 and Chattha *et al.*, 2012). During storage, seed quality can remain at the initial level or decline to a level that may make the seed unacceptable for planting purpose, what is related to many determinants: environment conditions during seed production, pests, diseases, seed oil content, seed moisture content, mechanical damages of seed in processing, storage longevity, packaging, pesticides, air temperature and relative air humidity in storage, biochemical injury of seed tissue (Al-Yahya, 2001; Šimic *et al.*, 2004; Guberac *et al.*, 2003; Heatherly and Elmore, 2004). Storage fungi i.e., *Aspergillus* spp. and *Penicillium* can grow in stored grain under bad storage conditions and cause serious losses (Mehrotra, 1983).

It is clear from the literature that *Aspergilli* are mainly storage fungi and the results given in table 8 are in support of this fact. As load of *Aspergilli* was found to be increased with increase in storage period and also with the type of storage containers. The storage containers like tin box and polythene bags supported maximum multiplication of *Aspergillus* population as compared to the same in gunny bags. Hence, it can be conducted that by using a proper storage containers population of *Aspergilli* can be controlled successfully without the aid of chemicals. Similarly, observations regarding use of chemicals as seed dressers for the control of *Aspergilli* are found to be interesting. The present study aimed to determine the effect of storage conditions in combination with packaging materials on the incidence of storage fungi.

MATERIALS AND METHODS:

Isolation of Aspergilli

1) Collection of Seed Samples:

The method described by Neergaard (1973) has been adopted for the collection of seed samples. Accordingly seed samples were collected from field, store houses and market places and from farmers. A composite sample was prepared by mixing the individual sample together, preserved in cloth bags at room temperature during the studies.

i) Detection and Identification of Seed Borne Fungi from Stored Plant Seeds by Blotter Plate Method:

A pair of white blotter paper of 8.5 cm diameter was jointly soaked in sterile distilled water, placed in pre-sterilized corning Petriplates of 10 cm diameter. Ten seeds per plate were placed at equal distance on the moist blotters. One hundred seeds were tested for each treatment. The plates were incubated at $25\pm2^{\circ}$ C under diurnal condition. On 7th day the seeds were examined under stereoscopic microscope for the preliminary determination of *Aspergillus*. Identification and further fungi occurred on seeds was made by preparing slides of the fungal growth and observing under compound microscope.

ii) Detection and Identification of Seed Borne Fungi by Agar Plate Method:

In this method, pre-sterilized corning glass Petri-plates of 10 cm diameter were poured with 25 ml of autoclaved water Agar (WA) medium. On cooling the medium, 10 seeds per plate were equispaced aseptically. One hundred seeds were tested for each treatment. The plates were incubated at 25 ± 2^{0} C under diurnal condition. On 7th day the seeds were examined under stereoscopic microscope for the preliminary determination of *Aspergillus*. Identification and further fungi occurred on seeds was made by preparing slides of the fungal growth and observing under compound microscope.

In order to isolate only internal seed mycoflora, seeds were pre-sterilized with 0.1% solution of mercuric chloride for 1 minute. Subsequently, thoroughly washed twice with sterile distilled water and placed on agar plates, blotter plate and water agar plates. Seeds without any such pre-treatments were employed for the total seed mycoflora (control). For blotter test, agar plate and water agar plate methods was followed as described by International Seed Testing Association, ISTA (1966) De Tempe (1970), Neergaard (1973) and Agarwal (1976).



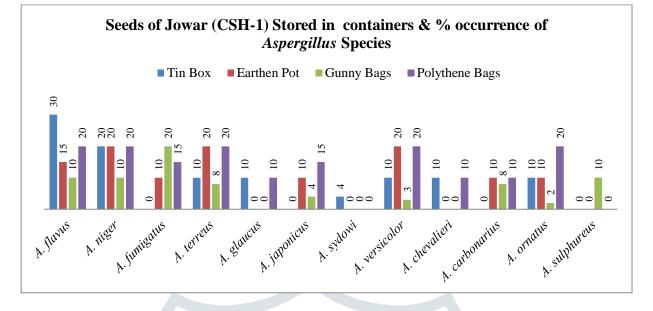
RESULTS AND DISCUSSIONS:

Table 1: Effect of Storage Containers on the Percent Incidence of Aspergillus Species

Sr. No.	Aspergillus Species Isolated	Seeds of Jowar (CSH-1) Stored in containers & %							
	Species Isolateu	occurrence of <i>Aspergillus</i> Species							
		Tin Box	Earthen Pot	Gunny	Polythene				
				Bags	Bags				
1	A. flavus	30	15	10	20				
2	A. niger	20	20	10	20				
3	A. fumigatus	-	10	20	15				
4	A. terreus	10	20	08	20				
5	A. glaucus	10		-	10				
6	A. japonicus	-	10	04	15				
7	A. sydowi	04	-		-				
8	A. versicolor	10	20	03	20				
9	A. chevalieri	10	-		10				
10	A. carbonarius		10	08	10				
11	A. ornatus	10	10	02	20				
12	A. sulphureus	-		10	-				

after 3 Months

Graph No 1: Effect of Different Storage Containers on Incidence of *Aspergillus* sp. After 3 months on Seeds of Jowar (CSH-1)

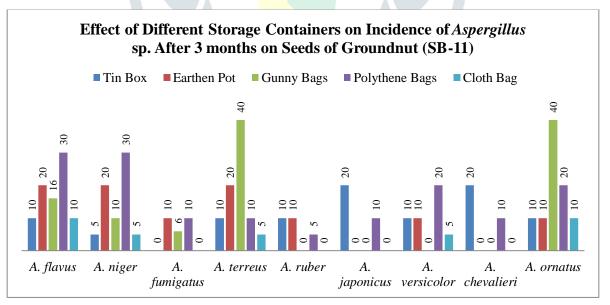




0	2019 JETIR	JETIR March 2019, Volume 6, Issue 3 Secus of Groundmut (SB-11) Stored in 2349-516							
		Species Isolated	Tin Box	Earthen	Gunny	Polythe	Cloth Bag		
				Pot	Bags	ne			
						Bags			
	1	A. flavus	10	20	16	30	10		
	2	A. niger	05	20	10	30	05		
	3	A. fumigatus	-	10	06	10	-		
	4	A. terreus	10	20	40	10	05		
	5	A. ruber	10	10	-	05	-		
	6	A. japonicus	20	-	-	10	-		
	7	A. versicolor	10	10	R -	20	05		
	8	A. chevalieri	20	-	-	10	-		
	9	A. ornatus	10	10	40	20	10		

Sr. No. Aspergillus Seeds of Groundnut (SB-11) Stored in ©

Graph- 2: Effect of Different Storage Containers on Incidence of Aspergillus sp. After 3 months on Seeds of Groundnut (SB-11)



It is clear from the results summarised in table 1 and graph 1 & 2 that the seeds of both the crops (jowar and groundnut) showed maximum incidence of Aspergillus species that were stored in the tin boxes followed by polythene bags while, the seeds stored in cloth bags

and gunny bags yielded less number of *Aspergillus* species. It is known from the literature that storage containers have their role in preventing or multiplying the original seed mycoflora in many cases. In order to confirm the same in case of different *Aspergillus* species, harvested mustard seeds of jowar and groundnut. The seeds were stored in irrespective containers at room temperature for the period of three months. After that the seeds were analysed for their load of *Aspergillus* species by using Potato Dextrose Agar plates.

A. *flavus* shows maximum incidence on seeds of jowar stored in tin box followed by polythene bags and whereas it was recorded that lowest percent incidence in gunny bags by *A. ruber*. A. sydowi was reported very low percent of incidence and only in tin box. No other containers supported growth of this species. A. sulphureus shows presence in lowest quantity on seeds of stored seeds of groundnut in gunny bags. The seeds stored in tin boxes and polythene bags favoured multiplication of *Aspergilli* more than the seeds stored in gunny bags.

REFERENCES:

- 1. Agarwal, V. K. (1976). Seed-borne fungi and viruses of some important crops. Research Bulletin 108 G. B. Pant Univ. of Agric. & Technol. Pantnagar.
- 2. Al-Yahya, S. A. 2001. Effect of storage conditions on germination in wheat. J. Agro. and Crop Sci. 186: 273- 279.
- Chattha, S. H., Jamali, L. A., Iibupoto, K. A. and Mangi, H. R. 2012.Effect of different packing materials and storage conditions on the viability of wheatseed (td-1 variety). Sci., Tech. and Dev. 31 (1): 10-18.
- 4. De Tempe, J. (1970). Testing cereal seeds for *Fusarium* infection in the Netherlands Pro. Int. Seed Test. Ass.35: 193-206.
- Guberac, V., Maric, S., Lalic, A., Drezner, G. and Zdunic, Z. 2003. Hermetically sealed storage of cereal seeds and its influence on vigor and germination. J. Agronomy and Crop Science 189: 54-56.
- Heatherly, L. G. and Elmore, R. W. 2004. Managing Inputs for Peak Production. Pages 451-536, in: Soybeans: Improvement, Production and Uses.Boerma H. R., Specht, J. E. (eds), 3rd Edition, Agronomy N-16, ASA, CSSA, SSSA, Madison, Wisconsin, USA.
- 7. ISTA (1966). International Seed Testing Association. Rules for seed health testing. 31: 1-152.
- Mehrotra, B. S. 1983. The impact of fungal infestation of cereal grains in field and storage. Pages 185-200, in: Recent Advances in Plant Pathology.Husain,A.,

Singh, K., Singh, B. P. and Agnihotri, V. P. (eds,), Lucknow Print House.

India.

- Neergaard P. (1973). Deterioration of seed borne pathogens by culture tests. Seed Sci. & Technol. 1: 217-254.
- Lacerda, A. D. S., Lazarini, E. and Filho, W. V. V. 2003. Storage of desiccated soybean seed and the evolution of physiologic, biochemical and sanitary characteristics. Braz. J. of Seeds 25: 97-10.
- 11. Šimic, B., Popovic, S. and Tucak, M. 2004. Influence of corn (*Zea mays* L.) inbred lines seed processing on their damage. Plant, Soil and Environment 50: 157-161.

